



# Meet Amblyseius swirskii (Acari: Phytoseiidae): a commonly used predatory mite in vegetable crops

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The predatory mite, *Amblyseius swirskii* Athias-Henriot (Acari: Phytoseiidae), is a generalist predator feeding on multiple soft-bodied insects and mite pest species. It is a biological control agent commercially available since 2005 that has become one of the top 3 most released biocontrol predators worldwide. It is commonly used to suppress whitefly populations (Hemiptera: Aleyrodidae), thrips (Thysanoptera), and spider mites (Acari: Tetranychidae) to a lesser degree. This predatory mite has been used as part of Integrated Pest Management (IPM) programs for vegetable (e.g., peppers, eggplants, cucumbers, squash) and ornamental (e.g., roses, chrysanthemums) crops in open fields and greenhouses, and some field crops such as cotton, but it has been demonstrated to be more successful at establishing reproductive populations and suppressing pests under protected structures. *Amblyseius swirskii* can feed on various food resources besides prey, including pollen or honeydew. It is successful at suppressing pests when used together with low-risk pesticides, when multiple prey are available, when prey and pollen are available naturally (neighboring flowering or companion plants present), or when pollen is supplemented in the field. This predator is a good option to control pests early in the season if shelter and food resources are available for its establishment.

Key words: biological control, predatory mite, natural enemy, Phytoseiidae

The predatory mite, *Amblyseius swirskii* Athias-Henriot (Acari: Phytoseiidae), belongs to the subfamily Amblyseiinae, the largest subfamily in the family Phytoseiidae with 1,478 nominal species (Chant and McMurtry 2007). This species is native to the Mediterranean region (Goleva and Zebitz 2013). Since it was commercially available in 2005, *A. swirskii* has been released in more than 50 countries in the world to feed on soft-bodied pests such as whiteflies, thrips, spider mites, and other pests of vegetables, fruits, and ornamental plants grown under protected structures (e.g., greenhouses, high tunnels) or in open fields (Nomikou et al. 2003, Calvo et al. 2015). The wide prey range of *A. swirskii* and its ability to feed on the pollen of various plant species make it a good predator for early-season establishment and is considered one of the most used predatory mites in the world.

# Life Cycle

The life cycle of phytoseiid mites, including *A. swirskii*, typically comprises 5 developmental stages: egg, larva, protonymph, deutonymph, and adult females and males. *Amblyseius swirskii* develops from egg to adult within a week at 25 °C and  $70\% \pm 5\%$  relative humidity (RH) and, unlike some pest mites (e.g., tetranychids), females must mate to produce eggs (Woolley 1988, Hoy 2011). Eggs are oval-shaped, translucent, and  $\sim 0.20 \times 0.15$  mm (Fig. 1). Eggs are usually laid on the underside of leaves individually or in clusters around the

intersection of the main and lateral veins. If present, females prefer to lay their eggs on non-glandular leaf trichomes or in clusters inside domatia (i.e., small pockets of nonglandular trichomes found commonly in some pepper plants and other plant species) to protect them from predators (Hoy 2011). Eggs last for approximately 1–2 days, during which time the egg develops into a 6-legged larva.

The larvae are small ( $\sim$ 0.22 × 0.16 mm), white to translucent in color, slow moving, and oval shaped (Fig. 2). This stage typically lasts for about 1–2 days. Larvae undergo one molt to develop into protonymphs that develop into deutonymphs after a second molt. Protonymphs and deutonymphs of *A. swirskii* have 8 legs, are orange colored, and are slightly larger than the larvae (Fig. 2). Each nymphal stage usually lasts for 1–2 days and feeds on insect pests such as first-instar thrips, first-instar whitefly immatures or whitefly eggs, and eggs or immature spider mites. After a short quiescent period, deutonymphs develop into adults (Hoy 2011, Calvo et al. 2015).

At this stage, adult females and males of *A. swirskii* are close to reaching their full size and are ready to mate. Adult females are about twice the size of the males and have pear-shaped bodies (Fig. 3). Adult females have a higher rate of predation to support the production of eggs. Adult males have oval bodies, usually move faster than females, and typically reach adulthood in a shorter time compared with *A. swirskii* females (Hoy 2011). The male guards a deutonymph female entering the quiescent stage and mates with the



**Fig. 1.** Two eggs of *Amblyseius swirskii* laid on the domatia of a bell pepper leaf accompanied by adult females (top) and cluster of eggs on the domatia of an ornamental pepper leaf (bottom).

female immediately after she molts into an adult by placing himself venter to venter with the female. Once in this position, the male transfers a sperm packet with his modified chelicerae to the female's spermatheca. One female can lay 1.5–2 eggs per day and between 30 and 40 eggs in her life span. Mature eggs occupy most of the female's body cavity, giving her a characteristic pear-shaped body when reaching full body size (Figs. 1 and 3) (Hoy 2011, Lopez and Liburd, unpublished data).

# **Optimal Environmental Conditions**

Amblyseius swirskii can develop and reproduce successfully over a range of temperatures from 15 to 36 °C, but its reproduction and survival are greatest at 25 °C. Adults can live for several weeks under favorable environmental conditions (22 ± 3 °C, 55%–85% RH), with access to shelter, water, and food resources (Abou-Awad et al. 2014, Al-Azzazy and Alhewairini 2020). Their survival, reproduction, and pest suppression ability are sensitive to temperature and RH and will decrease outside their optimal conditions (22–32 °C and 55–85% RH). It has been reported that A. swirskii population growth and development below 20 °C or above 38 °C could be slow and negatively impact their survival and ability to seek out prey (Ali and Zaher 2007, Lee and Gillespie 2011). Similarly, A. swirskii life stages are susceptible to changes in RH, resulting in long nymphal

developmental periods, low female fecundity, and low egg survival when RH is below 50%, compared with mite populations exposed to RH levels above 55% (Al-Azzazy and Alhewairini 2020, San et al. 2021). While nymphs and adults can drink water and feed on prey or plant exudates to compensate for water loss at low RH (<40%), eggs are susceptible to desiccation and reduced hatchability (San et al. 2021).

The surface of most host plant leaves has a unique microclimate thanks to gas interchange processes. The increased RH and temperatures on the surface of leaves can promote predatory mite survival when they are directly released on top of the plants. Similarly, enclosed release methods such as sachets create a microclimate with a higher RH, temperature, and water content to maintain *A. swirskii* optimal conditions for weeks and promote their establishment in the released crop (Solano-Rojas et al. 2022). Despite the advantages of microclimates within sachets or plants, it is vital for mites to have access to water. Moreover, the overall environmental conditions of the geographic location where these predatory mites are released must fall within the optimal ranges for *A. swirskii* to successfully establish and provide biocontrol services (San et al. 2021, Solano-Rojas et al. 2022).

Heavy rain events can also limit the establishment of *A. swirskii* in the open field. Thus, this predatory mite is most successfully used under protected structures where the desired average daily temperature throughout the year is between 20 and 30 °C and high RH is common (Lee and Gillespie 2011, Lopez and Liburd 2022).

# Mite Biology

## **Dispersal Capacity**

The ability of a predatory mite to suppress pests is strongly influenced by its capacity to disperse from the release site and seek prey throughout the crop (Pratt and Croft 2000, Buitenhuis et al. 2010, Parolin et al. 2014). Like most predatory mite species, A. swirskii disperses by walking through plants, soil, and planting material. They are highly mobile and can move from plant to plant using any bridges between the leaves (e.g., leaf bridges, pot surfaces, plastic or living mulch, and irrigation lines, to name a few) traveling at least 1 m within 24 h in high tunnel-grown pepper plants (Lopez et al. 2017) and at least 17 m within a week in open field-grown squash plants (Lopez and Liburd 2022). This is considered a high dispersal capacity for nonflying arthropods. Ambulatory dispersal is the main mode of dispersal for most predatory mites, followed by aerial dispersal. Amblyseius swirskii can disperse aerially, especially gravid females in search of prey, new oviposition sites, or alternative food sources. Females will not move far unless prey is scarce, or shelter is limited due to increasing predatory mite populations. Predatory mites can detect chemical cues released by the host plants, such as organic volatiles that indicate when the plant is infected by a pathogen (i.e., insect-transmitted pathogens), interacting with symbiotic microorganisms, or being fed on by herbivore arthropods (Schausberger et al. 2012). Similarly, they can estimate shelter and food availability based on encounters or interactions with other mites or based on chemical signals from other A. swirskii mites nearby or from other predatory mite species (e.g., webbing, molting sheds, among others) (Schausberger et al. 2012). These limiting situations may be disadvantageous for females' progeny; thus, females disperse to find new oviposition sites to secure food and shelter for their progeny and avoid cannibalism due to the increase in the population (Hoy 2011). When there is open space between plants, crops, or otherwise, females can use air currents to colonize new areas or



Fig. 2. Amblyseius swirskii larvae (top) and deutonymph (bottom).



Fig. 3. Adult female of Amblyseius swirskii.

crops. Because mites cannot decide where to fall but reach other places haphazardly, the presence of windbreakers, shrubs, and forest patches near neighboring crops plays an important role in their dispersal. Immature stages rarely move beyond one or a couple of leaves unless no food (prey or pollen) is available (Pratt and Croft 2000, Lopez et al. 2017). In addition, predatory mites can probably hitchhike on farm equipment, worker clothes, or greenhouse materials to colonize new plants or areas.

# **Promoting Establishment**

Companion planting or intercropping has been used as a diversification tactic mostly to promote biocontrol services by beneficial arthropods (Pratt and Croft 2000, Parolin et al. 2014). It involves growing other crop or noncrop plants with the cash crop. Insectary plants are often used as companion plants within vegetable crops to provide alternative shelter and food items to beneficial arthropods such as *A. swirskii*, promoting their establishment and dispersal toward the cropping system (Parolin et al. 2014). Examples of insectary plants used to encourage *A. swirskii* include sweet alyssum (Lobularia maritima (L.) Desv., Brassicales: Brassicaceae), buckwheat (Fagopyrum esculentum Moench, Caryophyllales: Polygonaceae), pinto bean (Phaseolus vulgaris L., Fabales: Fabaceae), sweet, chili, and ornamental peppers (Capsicum annum l., Solanaceae) (Fig. 4) (Xu and Enkegaard 2010, Calvo et al. 2011, Lopez et al. 2017).

#### **Food Resources**

Phytoseiid mites are classified into 4 groups based on their lifestyles (McMurtry et al. 2013). Amblyseius swirskii is categorized as a subtype III-b species consisting of mites that prefer to live on glabrous leaves (with little or no trichomes) and are generalist predators that feed on multiple insect and mite species. They can feed and reproduce on mites of families Eriophyidae (russet or gall mites), Tarsonemidae (e.g., broad mites), Tetranychidae (spider mites), Tenuipalpidae (false spider mites), and Tydeidae, as well as other soft-bodied organisms such as thrips (first-instar thrips), whiteflies (whitefly eggs and first and second instars), mealybugs, and nematodes, to name a few (McMurtry et al. 2013, Soleymani et al. 2016). In addition, they have been reported to feed and maintain themselves on microcrustacean species Artemia franciscana Kelogg (Crustacea: Artemiidae) as factitious food (i.e., a food source that the predator would not normally encounter in its natural habitat but on which the predatory mite can be reared for mass production) (Nguyen et al. 2013, Liburd et al. 2020).

Few reports of *A. swirskii* surviving on aphid honeydew exist, and it was reported as a poor food source for these predators (Ragusa and Swirski 1977). There are fewer reports of *A. swirskii* feeding on aphids. When *A. swirskii* was fed adult stages of *Aphis durantae* Theo (Hemiptera: Aphididae), long periods of nymphal development and low female fecundity were recorded indicating that these aphids were not an ideal food source for the mites (Ragusa and Swirski 1977, Ali and Zaher 2007).

As a generalist predatory mite, *A. swirskii* feeds not only on various prey but also on other food sources such as pollen, nectar, plant exudates, honeydew, and pycnial fluid from fungi in the absence of prey or as complementary food when prey is available (Nomikou et al. 2003, Goleva and Zebitz 2013). The combination of prey + alternative food resources (i.e., pollen) increases their reproductive capacity and the likelihood of establishment at early stages of the crop when pest abundance is low but flowering plants are present (McMurtry and Croft 1997, Calvo et al. 2015, Lopez et al. 2017) (Fig. 5). Therefore, supplementing pollen when releasing

A. swirskii could reduce or eliminate the need for multiple releases during the growing season and the cost minimized when pollen or multiple prey are present. Pollen subsidies and non-living factitious foods such as dry Artemia or Ephestia eggs can be purchased from multiple biological companies as a resource to promote predatory mite establishment, including A. swirskii.

No negative effects on biological control services have been reported associated with pollen supplementation for A. swirskii when prey is available. In contrast, pest control is improved when prey (e.g., twospotted spider mites, whiteflies, or thrips) is available, and pollen is supplemented, probably due to the increase in reproductive capacity caused by the presence of pollen (Nomikou et al. 2010, Schuldiner-Harpaz et al. 2016). Amblyseius swirskii populations can grow rapidly when fed only pollen, at a comparable rate to feeding on prey (Nomikou et al. 2010, Riahi et al. 2017). However, not all types of pollen are suitable for A. swirskii survival. Cattail (Typha spp.), pepper (Capsicum spp.), alyssum (Lobularia spp.), corn (Zea mays L.), and almond (Prunus dulcis Mill.) pollen are some of the pollen types that have been reported as suitable for development and population growth of A. swirskii (Henry et al. 2011, Schuldiner-Harpaz et al. 2016, Riahi et al. 2017). In contrast, some cucurbit pollen (i.e., squash pollen) does not seem like a preferred or even suitable food source for these mites (Lopez and Liburd 2022). Amblyseius swirskii has been reported feeding on extrafloral nectaries but their importance on population growth and survival is yet unknown (Henry et al. 2011).

# Distribution

Amblyseius swirskii is a predatory mite widely distributed in many parts of the world. It has been reported in Europe in many countries, including Spain, Italy, France, Greece, Turkey, Bulgaria, and the Netherlands. Amblyseius swirskii was originally described in 1962 from almond trees (P. dulcis) in Israel, where it naturally occurs on citrus, grapes, vegetables, and cotton (Swirski and Amitai 1997). In Spain, it is currently one of the most used predators in greenhouse crops, especially in pepper production, and it has also been found in citrus orchards and other outdoor crops (Bouagga et al. 2018). In Asia, A. swirskii has been reported in several countries, including Israel, Iran, and China. Similarly, this predatory mite has been reported in many African countries, including Egypt, South Africa, and Morocco. In all these countries, A. swirskii has been used as a biological control agent mostly in greenhouses and some open field crops for the control of whiteflies, thrips, and occasionally spider mites (Bouagga et al. 2018).

In North and South America, A. swirskii has been reported in the United States, Canada, Mexico, Brazil, and Chile. In the United States, it has been used in greenhouse crops, including tomatoes, cucumbers, and peppers, to control spider mites and thrips (Gerson et al. 2008, Calvo et al. 2015). Amblyseius swirskii is commonly found in Florida, where it was first discovered in the late 1990s and it is known to be established in many states since then, including California, Arizona, Georgia, Texas, Ohio, and Michigan (Gerson et al. 2008). The distribution of A. swirskii in the United States is influenced by various factors, including the introduction and spread of the mite by commercial growers for pest control purposes. However, despite its widespread use, the effectiveness of A. swirskii as a biological control agent can vary depending on the environmental conditions and the type of pest present (Gerson et al. 2008, Bouagga et al. 2018). Therefore, the distribution of this predatory mite may continue to evolve as vegetable, fruit, and ornamental growers seek effective pest control options that are sustainable and environmentally friendly.



Fig. 4. Adult females (bottom), immature (top), and egg (bottom) stages of Amblyseius swirskii in a bell pepper flower.

# **Economic Importance**

## Mass Rearing Methods

The most common methods of mass-rearing for phytoseiid mites around the world, including *A. swirskii*, are tri-trophic rearing systems. These consist of growing a plant host under greenhouses to rear prey, which will serve as food to sustain predatory mite populations. Commercial mass-rearing facilities often use bean plants as the foundation of their rearing systems due to the ease of cultivation and fast development into full-grown plants. Similarly, spider mites (*Tetranychus* spp., Tetranychidae) are typically chosen as prey because of their fast reproductive ability and population growth.

Most commercially available predatory mites (A. swirskii, Neoseiulus californicus, and N. cucumeris) are mass-reared only on factitious foods (i.e., a food source that the predator would not

normally encounter in its natural habitat) (Hoy 2011, Bouagga et al. 2018). The ability of *A. swirskii* to develop and reproduce factitious foods is a huge benefit in establishing mass-rearing systems (Barbosa and de Moraes 2015) at a relatively low cost. The mold mite *Tyrophagus putrescentiae* (Schrank) (Acari: Acaridae) and the dried fruit mite *Carpoglyphus lactis* L. (Acari: Carpoglyphidae) are factitious prey used successfully in the commercial rearing of *A. swirskii* around the world (Barbosa and de Moraes 2015, Pirayeshfar et al. 2020). Mass-produced *A. swirskii* are then distributed in various substrates consisting of bran, rice flakes, or both mixed with prey in bottle shakers, paper or plastic sachets, or releasing capsules.

Populations of A. swirskii can vary genetically among commercial biocontrol product providers. Because genetic variations are linked to resilience and predator performance when biocontrol



Fig. 5. Adult females of Amblyseius swirskii on ornamental pepper flower.

agents are released into crops, there may be performance benefits from using various providers when purchasing A. swirskii for biological control purposes (Paspati et al. 2019). In addition, the host plant and diet used to rear A. swirskii populations can significantly influence the behavior of the mites (Buitenhuis et al. 2014, Calvo et al. 2015). Various mass-reared predatory mites, including A. swirskii, have shown different performance levels (i.e., biocontrol services) based on the crop/plant where they are released and the pests they encounter. It has been demonstrated that releasing these commercial biocontrol agents in crops with different conditions from the ones they were reared on may harm their establishment and population growth (Buitenhuis et al. 2014). This is probably because the mites are accustomed to a plant structure and morphology type, diet, and environmental conditions different from those they encounter once released (Calvo and Knapp 2019). This is probably due to the predatory mite learning behavior when multiple generations have been reared on the same host plant or diet for long periods of time (Schausberger et al. 2020), as well as the species' genetically predetermined preferences. For instance, it is well known that A. swirskii avoids tomato and squash plants due to the glandular trichomes and may perform poorly in this crop or not establish at all. On the other hand, they thrive in pepper plants even when plants are not often used as rearing hosts in commercial rearing facilities. Therefore, early establishment of A. swirskii and/or pollen supplements, as well

as the use of flowering plants can be advantageous to enhance their performance (Xiao et al. 2012). Moreover, release rates of *A. swirskii* may need to be adjusted depending on the crop in which it is used (Buitenhuis et al. 2014).

## **Biocontrol Potential**

Amblyseius swirskii quickly became one of the most successful biocontrol agents after its introduction into the market in 2005 and is now released in more than 50 countries (Calvo et al. 2015, 2019). It is an effective predator of major pests found in vegetable production, such as the sweetpotato whitefly (Bemisia tabaci), the greenhouse whitefly (Trialeurodes vaporariorum Westwood, Hemiptera: Aleyrodidae), western flower thrips (Frankliniella occidentalis), melon thrips (Thrips palmi), and common blossom thrips (Frankliniella schultzei) (Xu and Enkegard 2010, McMurtry et al. 2013, Buitenhuis et al. 2015, Kutuk et al. 2016).

Amblyseius swirskii is well adapted to several vegetable crop hosts including cucumber, pepper, and eggplant (Nomikou et al. 2002, Stansly and Castillo 2009, Onzo et al. 2012, Farkas et al. 2016). This predatory mite effectively suppresses the sweetpotato whitefly and *E. occidentalis* populations in greenhouse-grown cucumber plants, suppressing up to 99% of whitefly nymphs compared to an exponential increase of more than 200 nymphs per leaf when

A. swirskii was absent (Calvo et al. 2011). Moreover, the presence of A. swirskii resulted in significant thrips suppression of more than 98% irrespective of the presence of whiteflies. The authors of this study also reported that predatory mite abundance was highest in the treatment combining A. swirskii, whitefly, and thrips. The option of targeting 2 pests with a single natural enemy has positive implications for biocontrol and resembles pest-predator complexes in field conditions (Messelink et al. 2010). In addition, it has been demonstrated that when multiple prey or food sources (prey + pollen) are available, A. swirskii survival is increased and populations are larger (Lopez et al. 2017).

In experiments to compare the effectiveness of several predatory mite species, Messelink et al. (2006) found that A. swirskii performed better suppressing F. occidentalis in cucumber compared with Euseius scutalis Athias-Henriot (Acari: Phytoseiidae) and N. cucumeris, the latter being the predatory mite commonly used for augmentative biological control of thrips in cucumber and other vegetable crops. In another study conducted on cucumber crops, Kakkar et al. (2016) demonstrated that A. swirskii provided effective control of T. palmi and F. schultzei under laboratory and semi-field conditions (shade house), and field trials. Their results demonstrated that A. swirskii may rival the effectiveness of chemical control strategies generally used for the management of these pests.

Few studies have reported the establishment of reproductive populations of A. swirskii in cucurbit crops such as squash (Kakkar et al. 2016, Lopez and Liburd 2022). This is probably because A. swirskii are more likely to establish in crops with glabrous leaves, such as peppers or cucumbers, than in leaves with abundant glandular trichomes such as some squash and tomato leaves (Xiao et al. 2012, Calvo et al. 2015). For instance, A. swirskii can be established in squash but prefers mature leaves that have fewer glandular trichomes. This predatory mite was rarely observed in younger leaves where these trichomes are most abundant (Lopez and Liburd 2022). Likewise, A. swirskii avoid tomato plants due to the presence of glandular trichomes consistent with this species' behavioral preference for glabrous leaves. The glandular trichomes on tomato leaves and stems not only limit A. swirskii movement but also have toxic effects on the nymphs and adult stages that are in contact with the secondary metabolites secreted in the tomato trichomes (Paspati et al. 2019). Thus, A. swirskii populations are often unsuccessful at establishing this crop.

Amblyseius swirskii has the ability to establish reproductive populations on other cucurbit crops besides cucumbers, such as summer squash in open fields and greenhouses. Companion plants serve as refugia by offering shelter, oviposition sites, and alternative food sources to support A. swirskii in times of prey (pest) scarcity (Xiao et al. 2012), given that this predatory mite is not likely to feed on squash pollen. Furthermore, parasitoids and larger predators that are also attracted to the companion plants, such as hoverflies (Syrphidae), big-eyed bugs (Geocoris spp.), and minute pirate bugs (Orius spp.), could feed on other pests in the system, such as aphids, while A. swirskii seeks out sweetpotato whiteflies and establishes earlier in the season by feeding on pollen or alternative prey.

In European countries, *A. swirskii* demonstrated its potential to suppress thrips and whiteflies in sweet peppers under protected structures in combination with *N. cucumeris*, and the minute pirate bug (*Orius laevigatus* Fieber, Hemiptera: Anthocoridae) (Van Maanen et al. 2010). *Amblyseius swirskii* use in sweet peppers expanded throughout the world soon after it was successfully used in Spain under greenhouse production (Van Maanen et al. 2010, Calvo et al. 2019). In addition, *A. swirskii* showed potential to be established in banker plant systems (i.e., open-rearing systems consisting of banker

plant, alternative food, and predatory mites used to maintain and disperse beneficial populations) and disperse from banker plants into pepper plants to successfully suppress populations of sweetpotato whiteflies, western flower thrips, and chilli thrips under laboratory and greenhouse conditions (Pratt and Croft 2000, Huang et al. 2011, Xiao et al. 2012). *Amblyseius swirskii* maintained low levels of the 3 pests for most of the experimental cycle (90 days) in laboratory conditions and more than 10 weeks under high tunnels (Lopez et al. 2017). Providing pollen (e.g., *Typha* pollen) to this predator may enhance its establishment and efficiency in controlling pests (Nomikou et al. 2010, Kutuk and Yigit 2011).

Additional studies have also reported good pest suppression by *A. swirskii* in onions and ornamentals such as roses and chrysanthemums (Onzo et al. 2012, Parolin et al. 2014, Calvo et al. 2015, Kumar et al. 2015). Similarly, cotton fields in Egypt have been reported as rich in *A. swirskii* individuals with abundant mites at the pleats of the lower surfaces of the leaves located in the middle part of cotton plants during the warm months of the season (Elshazly 2022).

## **Chemical Control Compatibility**

The effects of various insecticides used in vegetable production on A. swirskii have been evaluated. Most of the assessed products are limited to conventional cropping systems, including abamectin, metaflumizone, chlorantraniliprole, emamectin benzoate, fenpyroximate, methoxyfenozide, and flonicamid, to name a few (Colomer et al. 2011, Gradish et al. 2011, Amor et al. 2012, Lopez et al. 2015, Fernandez et al. 2017). The susceptibility of phytoseiid mites to plant protection products varies among developmental stages, with immature stages more susceptible to most pesticides than adult mites (Capinera 2008). Research has demonstrated that emamectin benzoate, milbectin, cyhexatin, fenpyroximate, and pyridaben have more negative impacts on phytoseiid adults and most also cause reduced fecundity (Amor et al. 2012, Lopez et al. 2015, Schmidt-Jeffris et al. 2021). While most also cause detrimental effects on immature stages, it has been shown that populations exposed to fenpyroximate treatments causing immature mortality below 50% can recover within a couple of weeks under optimal conditions (Lopez et al. 2015). Thus, the negative effects of even some harmful pesticides can be mitigated when well-timed applications are implemented.

Despite being considered a selective miticide in the past, studies testing the side effects of abamectin on phytoseiids have shown significant harm to predatory mites, and it is no longer considered selective (Gradish et al. 2011, Fernandez et al. 2017, Schmidt-Jeffris et al. 2021). Similarly, broad-spectrum pesticides such as etoxazole and most mitochondrial complex electron transport inhibitors (METI) are known to be harmful to predatory mites. However, some new METI pesticides such as bifenazate and acequinocyl are currently known as the least harmful products to predatory mites, together with hexythiazox, spiromesifen, and cyflumetofen (Fernandez et al. 2017, Schmidt-Jeffris et al. 2021), and some organic and reduce risk pesticides.

Exposure to reduce-risk pesticides such as potassium salts of fatty acids residues (e.g., M-Pede or Safer, also known as pesticidal soaps or soap salts) had minimal detrimental effects on all stages of the predatory mites (Lopez and Liburd, unpublished data). Reports of low *A. swirskii* mortality in M-Pede-treated squash may be related to the short persistence of its active ingredient. Similar findings were reported by Stanyard et al. (1998), who evaluated the toxicity of M-Pede on the predatory mite *Amblyseius fallacis* Garman (Acari: Phytoseiidae). This phytoseiid mite is used in apple trees for

the control of the European red mite (*Panonychus ulmi* Koch, Acari: Tetranychidae), and direct single applications of M-Pede showed little if any detrimental effects to the predatory mite. Potassium salts of fatty acids, as various other organic and/or reduce-risk pesticides, are known for their short persistence in the environment and often have a soil half-life of 1 day or less. However, when the frequency of M-Pede applications increased due to red mite outbreaks to more than 2 consecutive applications, *A. fallacis* populations were severely affected (Stanyard et al. 1998). Susceptibility to these potassium salts of fatty acids may vary depending on the predatory mite species.

Not all reduced-risk pesticides can be safely integrated with *A. swirskii* releases unless modifications on release schedules are enforced. For example, residues of azadirachtin + pyrethrins had detrimental effects on all stages of *A. swirskii*, with larvae being more negatively affected (49–73% mortality) than females and nymphs in laboratory experiments (Liburd et al. 2020). The pyrethrin component is a broad-spectrum chemical that targets the nervous system of insects and mites; it is very fast-acting and causes immediate paralysis. Azadirachtin has molting disruption properties (Yu 2008); thus, the larval stage was more susceptible to insecticide toxicity due to lower body sclerotization (Capinera 2008).

Substantially, high mortality for all A. swirskii developmental stages has been observed when mites were released 1-3 days after pesticide treatment (Liburd et al. 2020). Females and nymphs of the predatory mite released 1 day after treatment with reduced-risk insecticides (i.e., potassium salts of fatty acids and azadirachtin + pyrethrins) showed higher mortality rates when exposed to residues compared to the control (water sprays) in greenhouse-grown squash (Liburd et al. 2020). Releases of A. swirskii within 3 days after insecticide treatment should be avoided, and insecticide treatment should be scheduled at least a week after releasing A. swirskii to allow the predators to acclimate and establish in the crop. In addition, a deep understanding of the A. swirskii life cycle and population growth within the crop is imperative to schedule accompanying pesticide applications that minimize the negative effects of the predators present. Proper implementation of the plant protection product label and understanding its mode of action will also contribute to mitigating detrimental effects on natural enemies and improving pest suppression.

## **Concluding Remarks**

Amblyseius swirskii ranks in the top 3 of the most used commercially available predatory mites in the world. It is acclimated to multiple vegetable crops such as cucumbers, peppers, and eggplants, and can also be established in squash and cotton crops. It is known to prefer plant hosts with glabrous leaves or leaves that may offer specialized shelters like domatia. The presence of trichomes, particularly glandular trichomes, can limit the performance of some predatory mite species. The high density of glandular trichomes like the ones found on newly unfolded squash or tomato leaves are usually avoided by some generalist phytoseiids, including A. swirskii. Amblyseius swirskii is a generalist predatory mite species that can feed on a wide variety of insects and mites, but it is most successful at suppressing whiteflies, thrips, tarsonemids, and spider mites to a lesser degree. But it can also feed on a wide variety of alternative food items with some plant pollen at the top of the high-quality alternative foods comparable with some types of prey.

The main purpose of this article is not only to gather the most important aspects of *A. swirskii* biology but also to highlight the main components that must be considered for the successful use of *A. swirskii* in vegetable crops. These include, but are not limited to, the following:

- Identifying the pest or pests that need suppression and determining if *A. swirskii* is efficient at suppressing them.
- Considering if the plant morphology and fitness of the plant (e.g., presence of plant diseases, age of the crop, etc.) is suitable for the establishment and survival of A. swirskii.
- Examining environmental conditions within the target crop (e.g., temperature, RH, and water availability) to determine compatibility with the optimal conditions required for A. swirskii.
- Identifying trustworthy commercial providers where to acquire high-quality predatory mites. Guidelines to identify such providers are summarized by LeBeck and Leppla (2018) and more can be found at https://ipm.ifas.ufl.edu/applying/Biological\_Control.shtml.
- Gathering information regarding the developmental stage of the pests' populations in the crop to schedule well-timed predator releases and/or pesticide applications. The performance of *A. swirskii* can be promoted when small populations of pests are present and *A. swirskii* is used as a preventive tactic.
- Availability of pollen from crop or noncrop plants or pollen supplements can enhance their performance, as well as implementing other suppression practices in combination (e.g., cultural practices such as sanitation, isolation, etc.).
- If pest populations are already high before the introduction of biocontrol agents, the performance of *A. swirskii* can be limited, and pesticide applications may need to be scheduled prior to *A. swirskii* release.
- Choosing reduce-risk pesticides that mitigate detrimental effects to natural enemies for use with *A. swirskii* can promote the survival of these predators in the crop.
- Releasing A. swirskii more than 3 days after pesticide treatments is ideal to avoid detrimental effects due to pesticide residues. Similarly, pesticide applications should be conducted at least 1 wk after A. swirskii release to allow predator establishment.

Even though these general considerations could be followed for most phytoseiid mites, multiple factors may vary greatly among predatory mite species, including diet specificity, optimal environmental conditions, and preferred host plants, among others. *Amblyseius swirskii* is one of the most researched predatory mites in the world, together with *Phytoseiulus persimilis*, and this article is a basic overview of the knowledge related to *A. swirskii*.

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## **Author Contributions**

Lorena Lopez (Conceptualization [Lead], Investigation [Lead], Writing – original draft [Lead], Writing – review & editing [Lead])

## References

Abou-Awad BA, Hafez SM, Farahat BM. Bionomics and control of the broad mite *Polyphagotarsonemus latus* (Banks) (Acari: Tarsonemidae). Arch Phytopathol Pflanzenschutz. 2014;47(5):631–641.

Amor F, Medina P, Bengochea P, Canovas M, Vega P, Correia R, Garcia F, Gomez M, Budia F, Vinuela E, *et al.* Effect of emamectin benzoate under semi-field and field conditions on key predatory biological control agents

- used in vegetable greenhouses. Biocontrol Sci Technol. 2012;22(2):219–232. https://doi.org/10.1080/09583157.2011.650152
- Ali FS, Zaher MA. Effect of food and temperature on the biology of Typhlodrompis swirskii (Athias-Henriot) (Acari: Phytoseiidae). Acarines. 2007:1(1):17–21. https://doi.org/10.21608/ajesa.2007.4986
- Al-Azzazy MM, Alhewairini SS. Effect of temperature and humidity on development, reproduction, and predation rate of Amblyseius swirskii (Phytoseiidae) fed on Phyllocoptruta oleivora (Eriophyidae) and Eutetranychus orientalis (Tetranychidae). Int J Acarol. 2020:46(5):304–312. https://doi.org/10.1080/01647954.2020.1773922
- Barbosa MFC, de Moraes GJ. Evaluation of astigmatid mites as factitious food for rearing four predaceous phytoseiid mites (Acari: Astigmatina; Phytoseiidae). Biol Control. 2015;91:22–26.
- Bouagga S, Urbaneja A, Perez-Hedo M. Combined use of predatory mirids with Amblyseius swirskii (Acari: Phytoseiidae) to enhance pest management in sweet pepper. J Econ Entomol. 2018:111(3):1112–1120. https:// doi.org/10.1093/jee/toy072
- Buitenhuis R, Murphy G, Shipp L, Scott-Dupree C. Amblyseius swirskii in greenhouse production systems: a floricultural perspective. Exp Appl Acarol. 2015;65(4):451–464. https://doi.org/10.1007/s10493-014-9869-9
- Buitenhuis R, Shipp L, Scott-Dupree C, Brommit A, Lee W. Host plant effects on the behaviour and performance of Amblyseius swirskii (Acari: Phytoseiidae). Exp Appl Acarol. 2014;62:171–180. https://doi.org/10.1007/s10493-013-9735-1
- Buitenhuis R, Shipp L, Scott-Dupree C. Intra-guild vs extra-guild prey: effect on predator fitness and preference of Amblyseius swirskii (Athias-Henriot) and Neoseiulus cucumeris (Oudemans) (Acari: Phytoseiidae). Bull Entomol Res. 2010:100(2):167–173. https://doi.org/10.1017/S0007485309006944
- Calvo FJ, Knapp M. Biological control of insect pests using predators and parasitoids. In: Omkar M, editor. Amblyseius swirskii: a generalist phytoseiid predator with potential for the control of whitefly, thrips, and spider mites. Singapore: Springer; 2019. p. 231–254.
- Calvo FJ, Bolckmans K, Belda JE. Control of Bemisia tabaci and Frankliniella occidentalis in cucumber by Amblyseius swirskii. Biocontrol. 2011:56(2):185–192. https://doi.org/10.1007/s10526-010-9319-5
- Calvo FJ, Knapp M, van Houten YM, Hoogerbrugge H, Belda JE. *Amblyseius swirskii*: what made this predatory mite such a successful biocontrol agent? Exp Appl Acarol. 2015:65(4):419–433.
- Capinera JL. Encyclopedia of entomology. 2nd ed. Berlin (Germany): Springer Science; 2008.
- Chant DA, McMurtry JA. Illustrated keys and diagnoses for the genera and subgenera of the Phytoseiidae of the world (Acari: Mesostigmata). West Bloomfield (MI): Indira Publishing House; 2007.
- Colomer I, Aguado P, Medina P, Heredia MR, Fereres A, Belda EJ, Vinuela E. Field trial measuring the compatibility of methoxyfenozide and flonicamid with *Orius laevigatus* Fieber (Hemiptera: Anthocoridae) and *Amblyseius swirskii* (Athias-Henriot) (Acari: Phytoseiidae) in a commercial pepper greenhouse. Pest Manag Sci. 2011:67:1237–1244.
- Elshazly MMY. Obtaining large numbers of predatory mites, namely *Amblyseius swirskii* Athias-Henriot (Acari: Phytoseiidae), from cotton fields rich in it, and accumulating them on a few leaves. 2022 [accessed 2023 May 8]. https://www.researchsquare.com/article/rs-1358249/v1.
- Farkas P, Bagi N, Szabó Á, Ladányi M, Kis K, Sojnóczki A, Reiter D, Pénzes B, Fail J. Biological control of thrips pests (Thysanoptera: Thripidae) in a commercial greenhouse in Hungary. Pol J Entomol. 2016:85(4):437–451.
- Fernandez MM, Medina P, Wanumen A, Estal P, Smagghe G, Viñuela E. Compatibility of sulfoxaflor and other modern pesticides with adults of the predatory mite *Amblyseius swirskii*. Residual contact and persistence studies. BioControl. 2017:62:107–208. https://doi.org/10.1007/s10526-017-9784-1
- Gradish A, Scott-Dupree E, Shipp CD, Harris L, Ferguson CR. Effect of reduced risk pesticides on greenhouse vegetable arthropod biological control agents. Pest Manag Sci. 2011:67(1):82–86.
- Gerson U, Smiley RL, Ochoa R. Mites (Acari) for pest control. New York: John Wiley & Sons; 2008.
- Goleva I, Zebitz CP. Suitability of different pollen as alternative food for the predatory mite Amblyseius swirskii (Acari, Phytoseiidae). Exp Appl Acarol. 2013:61(3):259–283. https://doi.org/10.1007/s10493-013-9700-z

- Henry ME, Brown SP, Arthurs S, Osborne LS. Evaluation of ten groundcovers as a landscape banker plant for Amblyseius swirskii. Proc Fla State Hort Soc 2011:124:317–320
- Hoy MA. Agricultural acarology: introduction to integrated mite management. 1st ed. CRC Press, Boca Raton, FL. 2011.
- Huang N, Enkegaard A, Osborne LS, Ramakers PMJ, Messelink GJ, Pijnakker J, Murphy G. The banker plant method in biological control. Crit Rev Plant Sci. 2011:30(3):259–278. https://doi.org/10.1080/07352689.2011. 572055
- Kakkar G, Kumar V, Seal DR, Liburd OE, Stansly PA. Predation by Neoseiulus cucumeris and Amblyseius swirskii on Thrips palmi and Frankliniella schultzei on cucumber. Biol Control. 2016:92:85–91. https://doi. org/10.1016/j.biocontrol.2015.10.004
- Kumar V, Xiao Y, McKenzie CL, Osborne LS. Early establishment of the phytoseiid *Amblyseius swirskii* (Acari: Phytoseiidae) on pepper seedlings in a predator-in-first approach. Exp Appl Acarol. 2015:65(4):465–481. https://doi.org/10.1007/s10493-015-9895-2
- Kutuk H, Karacaoglu M, Tufekli M, Villanueva RT. Failure of biological control of Frankliniella occidentalis on protected eggplants using Amblyseius swirskii in the Mediterranean region of turkey. Turk J Agric For. 2016;40(1):13–17.
- Kutuk H, Yigit A. Pre-establishment of Amblyseius swirskii (Athias-Henriot) (Acari: Phytoseiidae) using Pinus brutia (Ten.) (Pinales: Pinaceae) pollen for thrips (Thysanoptera: Thripidae) control in greenhouse peppers. Int J Acarol. 2011;37(1):95–101.
- LeBeck LM, Leppla NC. Guidelines for purchasing and using commercial natural enemies in North America. 2018 [accessed 2023 July 31]. https://ipm.ifas.ufl.edu/applying/Revised%20Guidelines%20for%20Natural%20Enemies%20to%20Post%20120720%20(%20042321%20Update).pdf.
- Lee HS, Gillespie DR. Life tables and development of *Amblyseius swirskii* (Acari: Phytoseiidae) at different temperatures. Exp Appl Acarol. 2011;53(1):17–27. https://doi.org/10.1007/s10493-010-9385-5
- Liburd OE, Lopez L, Carrillo D, Revynthi AM, Olaniyi O, Akyazi R. Integrated management of insect pests: current and future developments. In: Kogan M, Heinrichs E, editors. Integrated pest management of mites. Cambridge (UK): Burleigh Dodds Science Publishing; 2020.
- Lopez L, Liburd OE. Can the introduction of companion plants increase biological control services of key pests in organic squash?. Exp Appl Entomol. 2022;170(5):402–418. https://doi.org/10.1111/eea.13147
- Lopez L, Smith HA, Hoy MA, Bloomquist JR. Acute toxicity and sublethal effects of fenpyroximate to Amblyseius swirskii (Acari: Phytoseiidae). J Econ Entomol. 2015:108(3):1047–1053. https://doi.org/10.1093/jee/tov033
- Lopez L, Smith HA, Hoy MA, Cave RD. Dispersal of Amblyseius swirskii (Acari: Phytoseiidae) on high-tunnel bell peppers in presence or absence of Polyphagotarsonemus latus (Acari: Tarsonemidae). J Insect Sci. 2017;17(1):1–7.
- McMurtry JA, de Moraes GJ, Sourassou NF. Revision of the lifestyles of phytoseiid mites (Acari: Phytoseiidae) and implications for biological control strategies. Syst Appl Acarol. 2013:18(4):297–320.
- McMurtry JA, Croft BA. Life-styles of Phytoseiid mites and their roles in biological control. Annu Rev Entomol. 1997:42:291–321. https://doi. org/10.1146/annurev.ento.42.1.291
- Messelink GJ, Van Maanen R, Van Holstein-Saj R, Sabelis MW, Janssen A. Pest species diversity enhances control of spider mites and whiteflies by a generalist phytoseiid predator. Biocontrol. 2010:55(3):387–398. https://doi.org/10.1007/s10526-009-9258-1
- Messelink GJ, Van Steenpaal SEF, Ramakers PMJ. Evaluation of phytoseiid predators for control of western flower thrips on greenhouse cucumber. Biocontrol. 2006:51(6):753–768. https://doi.org/10.1007/s10526-006-9013-9
- Nomikou M, Sabelis MW, Janssen A. Pollen subsidies promote whitefly control through the numerical response of predatory mites. BioControl. 2010:55:253–260.
- Nomikou M, Janssen A, Sabelis MW. Phytoseiid predators of whiteflies feed and reproduce on non-prey food sources. Exp Appl Acarol. 2003;31:15–26. https://doi.org/10.1023/B;APPA.0000005142.31959.e8
- Nomikou M, Janssen A, Schraag R, Sabelis MW. Phytoseiid predators suppress populations of *Bemisia tabaci* on cucumber plants with alternative food. Exp Appl Entomol. 2002:27(1-2):57–68.

- Nguyen DT, Vangansbeke D, Lü X, De Clercq P. Development and reproduction of the predatory mite *Amblyseius swirskii* on artificial diets. BioControl. 2013;58(3):369–377. https://doi.org/10.1007/s10526-012-9502-y
- Onzo A, Houedokoho AF, Hanna R. Potential of the predatory mite, Amblyseius swirskii to suppress the broad mite, Polyphagotarsonemus latus on the gboma eggplant, Solanum macrocarpon. J Insect Sci. 2012;12(7):1–11, https://doi.org/10.1673/031.012.0701
- Parolin P, Bresch C, Poncet C, Desneux N. Introducing the term 'Biocontrol Plants' for integrated pest management. Sci Agric. 2014;71(1):77–80. https://doi.org/10.1590/s0103-90162014000100011
- Paspati A, Ferguson KB, Verhulst EC, Urbaneja A, González-Cabrera J, Pannebakker BA. Effect of mass rearing on the genetic diversity of the predatory mite *Amblyseius swirskii*. Entomol Exp Appl. 2019:167:670– 681. https://doi.org/10.1111/eea.12811
- Paspati A, Rambla JL, López Gresa MP, Arbona V, Gómez-Cadenas A, Granell A, González-Cabrera J, Urbaneja A. Tomato trichomes are deadly hurdles limiting the establishment of *Amblyseius swirskii* Athias-Henriot (Acari: Phytoseiidae). Biol Control. 2019:157:104572. https://doi.org/10.1016/j.biocontrol.2021.104572
- Pirayeshfar F, Safavi SA, Moayeri HRS, Messelink GJ. The potential of highly nutritious frozen stages of *Tyrophagus putrescentiae* as a supplemental food source for the predatory mite *Amblyseius swirskii*. Biocontrol Sci Technol. 2020:30(5):403–417.
- Pratt PD, Croft BA. Banker plants: evaluation of release strategies for predatory mites. J Environ Hort. 2000:18(4):211–217. https://doi. org/10.24266/0738-2898-18.4.211
- Ragusa S, Swirski E. Feeding habits, post-embryonic and adult survival, mating, virility and fecundity of the predacious mite *Amblyseius swirskii* [Acarina: Phytoseiidae] on some coccids and mealybugs. Entomophaga. 1977:22(4):383–392. https://doi.org/10.1007/bf02373263
- Riahi E, Fathipour Y, Talebi AA, Mehrabadi M. Linking life table and consumption rate of *Amblyseius swirskii* (Acari: Phytoseiidae) in presence and absence of different pollens. Ann Entomol Soc Am. 2017:110(2):244–253. https://doi.org/10.1093/aesa/saw091
- San PP, Tuda M, Takagi M. Impact of relative humidity and water availability on the life history of the predatory mite *Amblyseius swirskii*. BioControl. 2021:66(4):497–510. https://doi.org/10.1007/s10526-021-10081-y
- Schausberger P, Seiter M, Raspotnig G. Innate and learned responses of foraging predatory mites to polar and non-polar fractions of thrips' chemical cues. Biol Control. 2020:151:104371. https://doi.org/10.1016/j. biocontrol.2020.104371
- Schausberger P, Peneder S, Jürschik S, Hoffmann D. Mycorrhiza changes plant volatiles to attract spider mite enemies. Funct Ecol. 2012;26(2):441–449. https://doi.org/10.1111/j.1365-2435.2011.01947.x

- Schmidt-Jeffris RA, Beers EH, Sater C. Meta-analysis and review of pesticide non-target effects on phytoseiids, key biological control agents. Pest Manag Sci. 2021:77(11):4848–4862. https://doi.org/10.1002/ps.6531
- Schuldiner-Harpaz T, Coll M, Weintraub PG. Prey and pollen food choice depends on previous diet in an omnivorous predatory mite. Environ Entomol. 2016:45(4):995–998. https://doi.org/10.1093/ee/ nvw063
- Solano-Rojas Y, Gallego JR, Gamez M, Lopez I, Castillo P, Cabello T. Effect of relative humidity on the population dynamics of the predator Amblyseius swirskii and its prey Carpoglyphus lactis in the context of slow-release sachets for use in biological control in greenhouses. Plants. 2022:11(19):2493–2112. https://doi.org/10.3390/plants11192493
- Soleymani S, Hakimitabar M, Seiedy M. Prey preference of predatory mite Amblyseius swirskii (Acari: Phytoseiidae) on Tetranychus urticae (Acari: Tetranychidae) and Bemisia tabaci (Hemiptera: Aleyrodidae). Biocontrol Sci Technol. 2016:26(4):562–569. https://doi.org/10.1080/09583157.201 5 1133808
- Stansly PA, Castillo J. Control of broad mites, Spider mites, and Whiteflies using predaceous mites in open-field pepper and eggplant. Proc Fla State Hort Soc. 2009:122:253–257.
- Stanyard MJ, Fosteh RE, Gibb TJ. Population dynamics of Amblyseius fallacis (Acari: Phytoseiidae) and European red mite (Acari: Tetranychidae) in apple trees treated with selected acaricides. J Econ Entomol. 1998:91(1):217–225.
- Swirski E, Amitai S. Annotated list of Phytoseiid mites (Mesostigmata: Phytoseiidae) in Israel. Isr J Entomol. 1997:31:21–46.
- Van Maanen R, Vila E, Sabelis MW, Janssen A. Biological control of broad mites (*Polyphagotarsonemus latus*) with the generalist predator *Amblyseius swirskii*. Exp Appl Acarol. 2010:52(1):29–34. https://doi. org/10.1007/s10493-010-9343-2
- Woolley TA. Acarology: mites and human welfare. New York (NY): John Wiley & Sons, Inc; 1988.
- Xiao Y, Avery P, Chen J, McKenzie C, Osborne L. Ornamental peppers as banker plants for establishment of *Amblyseius swirskii* (Acari: Phytoseiidae) for biological control of multiple pests in greenhouse vegetable production. Biol Control. 2012;63(3):279–286. https://doi.org/10.1016/j.biocontrol.2012.09.007
- Xu X, Enkegaard A. Prey preference of the predatory mite, Amblyseius swirskii between first instar western flower thrips Frankliniella occidentalis and nymphs of the twospotted spider mite Tetranychus urticae. J Insect Sci. 2010:10(149):1–11. https://doi.org/10.1673/031.010.14109
- Yu SJ. The toxicology and biochemistry of insecticides. 1st ed. CRC Press/ Taylor & Francis, Boca Raton, FL. 2008. p. 296.